



Nigerian medicinal plants with *in vivo* anti diabetic activity: A systematic review and Meta-analysis

Ahmed HA^{1*}, Mahmud AF², M Lawal³, Alkali YI⁴

^{1,3} Department of Pharmacognosy and Ethnopharmacy, Faculty of Pharmaceutical Sciences, Usmanu Danfodiyo University, Sokoto, Nigeria

² Department of Pharmacognosy and Drug Development, Faculty of Pharmaceutical Sciences, Ahmadu Bello University, Zaria, Nigeria

⁴ Department of Pharmacology and Toxicology, Usmanu Danfodiyo University, Sokoto, Nigeria

DOI: <https://doi.org/10.33545/26646862.2019.v1.i2a.9>

Abstract

The aim of this study was to systematically review the literature for scientific information on Nigerian medicinal plants with antidiabetic activity providing researchers with valuable information on the most common plants species and methods used. A search was carried out for studies investigating the antidiabetic activity of Nigerian medicinal plants using the following scientific data bases; science direct, Google scholar, PubMed and Directory of open access journals (DOAJ). The search included articles from January 2014-2019. To assess the quality of the Studies, the Jadad scale was used. The information recorded included; plant name, family, diabetic model used, Intervention, duration of study and Outcome. Randomized controlled trials which evaluated the hypoglycemic effects of plants with significant outcome of decrease in fasting blood glucose (FBG) were included. Most of the plants in this study showed significant glycemic control. There is need for further studies to identify the phytochemicals responsible for the activity.

Keywords: medicinal plants, antidiabetic, systematic review, meta-analysis

1. Introduction

Worldwide, approximately 422 million people suffer from Diabetes mellitus ^[1] and by the year 2040, the number of people with Diabetes mellitus is estimated to rise to 642 million globally ^[2]. Diabetes Mellitus (DM) is a chronic metabolic disorder characterized by inadequate secretion of insulin or development of insulin resistance. The disorder is characterized by hyperglycemia, polyuria, polydipsia and polyphagia. Diabetes mellitus is classified into two main types namely; type 1 and type 2 diabetes mellitus. Type 1 which is also called as insulin dependent diabetes is an autoimmune disorder leading to destruction of pancreatic β -cells which leads to decrease in the secretion of insulin ^[3]. The increase in prevalence of diabetes and the side-effects of current drugs used in management of diabetes mellitus necessitates the need to search for alternatives ^[4,5]. About 80% of the populations in developing countries rely on medicinal plants as the main source of treatment of diseases ^[6, 7, 8].

The catalogue of antidiabetic medicinal plants is growing at a high rate especially in Africa. This may be due to the high cost of managing diabetes which forces patients to source for alternative and cheaper options. This overreliance on antidiabetic medicinal plants has probably invoked scientists to bioassay these plants in an effort to identify more hypoglycemic medicinal plants. The antidiabetic potential of some medicinal plants extracts has been demonstrated in human and animal models of type II diabetes ^[9]. There is an increase in the number of scientific publications on medicinal plants and diabetes ^[10]. Therefore the aim of this study is to systematically review the literature for scientific information on Nigerian medicinal plants with antidiabetic activity providing researchers with valuable information on the most common plants species and methods used.

2. Materials and Method

2.1 Sources of information and eligibility criteria

The present meta-analysis was reported according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines ^[11]. A search was carried out for studies investigating the antidiabetic activity of Nigerian medicinal plants using the following scientific data bases; science direct, Google scholar, Science direct, PubMed, Hindawi and Directory of open access journals (DOAJ). The search included articles from January 2014-2019. The Jadad scale was used to evaluate the quality of the studies ^[12]. The search terms were antidiabetic activity, medicinal plants. The search was limited to Nigerian authors. The information recorded include; name of plant, family, local name(s), diabetic model used, dose used, part of plant used, duration of study and Outcome.

2.2 Inclusion Criteria

Randomized controlled trials which evaluated the antidiabetic effects of plants with significant outcome of decrease in blood glucose were included. The search was limited to articles in English language and authors with Nigerian affiliation.

2.3 Exclusion Criteria

Articles with Jadad score less than three (3), studies targeting specific mechanism of action, *in vitro studies* and studies on diabetic complications were excluded.

3. Results

A total of 15,283 records were identified through database search; 3,830 entries for PubMed, 9591 from science direct, 121 from

Table 1: Animal studies on Nigerian Medicinal Plants

S/N	Plants	Target	Groups	Intervention	Duration	Drug standard	Author(s)
1	<i>Tamarindus Indica</i>	Alloxan	Diabetic rats	1000mg/kg 90% methanol stem-bark extract	24hours	Metformin	Yerima, 2018 ^[13]
2	<i>Clerodendrum volubile</i>	Streptozocin	normal and diabetic rats	400mg/kg n-butanol, ethyl acetate, methanol leaf extracts	21days	glibenclamide	Erukainure <i>et al.</i> , 2018 ^[14]
3	<i>Denntia tripetala</i>	Streptozocin	normal and diabetic rats	50mg/kg seed methanol extract	10days	glibenclamide	Innocent <i>et al.</i> , 2017 ^[15]
4	<i>Ocimum canum</i>	Streptozocin	diabetic rats	100mg/kg methanol leaf extract	15days	glibenclamide	Ononamadu <i>et al.</i> , 2019 ^[16]
5	<i>Gouania longipetala</i>	Alloxan	diabetic rats	150mg/kg methanol leaf extract	21days	glibenclamide	Maxwell <i>et al.</i> , 2014 ^[17]
6	<i>Thaumatococcus danielli</i>	Streptozocin	Normal and diabetic rats	500mg/kg ethanol leaf extract	14days	Glibenclamide	Folorunsho <i>et al.</i> , 2019 ^[18]
7	<i>Balanites aegyptiaca</i>	Streptozocin	Normal and diabetic	200mg/kg 70% ethanol stem-bark extract	120minutes	metformin	Mhya <i>et al.</i> , 2016 ^[19]
8	<i>Cucumis sativus</i>	Alloxan	Diabetic rats	400mg/kg Aqueous stem-bark extract	15days	metformin	Ibitoye <i>et al.</i> , 2017 ^[20]
9	<i>Morinda lucida</i>	Alloxan	Normal and diabetic rats	500mg/kg stem-bark aqueous extract	8days	glibenclamide	Adeneye <i>et al.</i> , 2017 ^[21]
10	<i>Ocimum gratissimum</i>	Streptozocin	Normal and diabetic rats	250mg/kg 80% methanol leaf extract	28days	metformin	Stanley <i>et al.</i> , 2017 ^[22]
11	<i>Sapium ellipticum</i>	Streptozocin	Normal and diabetic rats	800mg/kg ethanol	21days	metformin	Osasaneng 2017 ^[23]
12	<i>Citrus aurantifolia</i>	Alloxan	normal and diabetic rats	100mg/kg Aqueous	14 days	Metformin	Ibrahim <i>et al.</i> , 2018 ^[24]
13	<i>Ricinus communis</i>	alloxan	normal and diabetic rats	400mg/kg Seed methanol extract	21 days	metformin	Nwodo <i>et al.</i> , 2016 ^[25]
14	<i>Salacia lehmbachii</i>	alloxan	normal and diabetic rats	400mg/kg 96% methanol Stem bark	21 days	glibenclamide	Joseph <i>et al.</i> , 2018 ^[26]
15	<i>Uvaria chamae</i>	Alloxan		400 mg/kg Root ethanol	15days	glibenclamide and pioglitazone	Jonathan <i>et al.</i> , 2018 ^[27]
16	<i>Aristolochia ringens</i>	streptozotocin	diabetic rats	150mg/kg Methanol leaf and stem bark	7days	metformin	Sulayman <i>et al.</i> , 2019 ^[28]
17	<i>Jatropha curcas</i>	Alloxan	normal and diabetic rats	Methanol leaf	7days	metformin	Aina <i>et al.</i> , 2016 ^[29]
18	<i>Massularia acuminata</i>	alloxan	diabetic rats	89.44 mg/kg 70% ethanol Stem	5days	glibenclamide	Emmanuel <i>et al.</i> , 2018 ^[30]
19	<i>Strophanthus hispidus</i>	Alloxan	normal and diabetic rats	200mg/kg Aqueous Root	28 days	Glibenclamide	Fageyinbo <i>et al.</i> , 2019 ^[31]
20	<i>Moringa oleifera</i>	Alloxan	diabetic rats	Ethanol 800mg/kg seeds, leaves, stem, roots	6hours	acarbose	Idakwoji <i>et al.</i> , 2016 ^[32]
21	<i>Vernonia amygdalina</i>	streptozotocin	diabetic rats	200mg/kg Chloroform Leaves	28days	metformin	Stanley <i>et al.</i> , 2019 ^[33]
22	<i>Anogeissus leiocarpus</i>	alloxan	diabetic rats	200mg/kg Ethanol roots	24hours	glibenclamide	Omeje <i>et al.</i> , 2016 ^[34]
23	<i>Mitracarpus Scabrum</i>	Alloxan	normal and diabetic rats	300mg/kg leaves	14days	glibenclamide	Abubakar <i>et al.</i> , 2016 ^[35]
24	<i>Hoslundia opposita</i>	alloxan	normal and diabetic rats	220 mg/kg Seed oil	96hours	metformin	Jubril, <i>et al.</i> , 2014 ^[36]
25	<i>Senna singueana</i>	Streptozotocin	normal and diabetic rats	300mg/kg Acetone Stem-bark	28days	metformin	Mohammed and Shahidul 2014 ^[37]
26	<i>Parkia biglobosa</i>	streptozotocin	normal and diabetic rats	300mg/kg n-butanol leaves	5days	Metformin	Mohammed <i>et al.</i> , 2016 ^[38]
27	<i>Blighia sapida</i>	Alloxan	normal and diabetic rats	400mg/kg methanol leaves	28days	glibenclamide	Oloyede <i>et al.</i> , 2014 ^[39]
28	<i>Chrysophyllum albidum</i>	alloxan	normal and diabetic rats	200 mg/kg ethanol Seeds	7days	metformin	Engwa <i>et al.</i> , 2016 ^[40]
29	<i>Anthocleista vogelii</i>	alloxan	normal and diabetic rats	200mg/kg Root bark Chloroform	21days	glibenclamide	Anyanwu <i>et al.</i> , 2018 ^[41]
30	<i>Synsepalum dulcificum</i>	streptozotocin	normal and diabetic rats	60 mg/kg 80% methanol Leaves	21days	glibenclamide	Obafemi, <i>et al.</i> , 2019 ^[42]
31	<i>Cola nitida</i>	fructose and streptozotocin	normal and diabetic rats	300 mg/kg aqueous seeds	42days	Metformin	Ochuko <i>et al</i> 2019 ^[43]
32	<i>Curculigo pilosa</i>	streptozotocin	normal and diabetic rats	methanol, n-butanol Rhizomes	28days	glibenclamide	Kayode and Charles, 2019 ^[44]
33	<i>Phyllanthus fraternus</i>	streptozotocin	normal and diabetic rats	200-300mg/kg leaves	28days	metformin	Nadro and Elkanah 2017 ^[45]
34	<i>Phoenix Dactylifera</i>	Alloxan	normal and diabetic rats	400mg/kg ethanol seeds	14days	glibenclamide	Abiola <i>et al.</i> , 2018 ^[46]
35	<i>Acacia ataxacantha</i>	streptozotocin	normal and diabetic rats	125-500mg/kg 95% ethanol Roots	7days	metformin	Arise <i>et al.</i> , 2016 ^[47]
36	<i>Borassus aethiopum</i>	Alloxan	normal and diabetic rats	100-500mg/kg aqueous fruit	28days	glibenclamide	Issaka <i>et al.</i> , 2016 ^[48]
37	<i>Cnidioscolus aconitifolius</i>	streptozotocin	normal and diabetic rats	500 mg/kg	28days	glibenclamide	Achi <i>et al.</i> , 2016 ^[49]
38	<i>Adansonia digitata</i>	Alloxan	normal and diabetic rats	200mg/kg fruit aqueous	14days	Chlorpropamide	Muhammad <i>et al.</i> , 2016 ^[50]
39	<i>Newbouldia laevis</i>	alloxan	normal and diabetic rats	250-1000mg/kg Hydro alcohol leaves	24hours	glibenclamide	Osigwe <i>et al.</i> , 2015 ^[51]
40	<i>Ceiba pentandra</i>	Alloxan	normal and diabetic rats	200-800mg/kg stem-bark methanol	14days	glibenclamide	Odoh <i>et al.</i> , 2016 ^[52]
41	<i>Tabernaemontana pachysiphon</i>	alloxan	normal and diabetic rats	125-1000mg/kg methanol stem bark	14days	glibenclamide	Osamuyi <i>et al.</i> , 2017 ^[53]
42	<i>Ageratum conyzoides</i>	streptozotocin	normal and diabetic rats	200-800mg/kg ethyl acetate, leaves	14days	metformin	Agbafor <i>et al.</i> , 2015 ^[54]
43	<i>Myrianthus arboreus</i>	streptozotocin	normal and diabetic rats	100-400mg/kg Ethanol stem bark	28days	glibenclamide	Dickson <i>et al.</i> , 2016 ^[55]
44	<i>Dioscoreophyllum cumminsii</i>	alloxan	diabetic rats	50-300mg/kg leaves	21days	glibenclamide	Oloyede <i>et al.</i> , 2015 ^[56]

Directory of open access journals (DOAJ), 1051 from Hindawi and 690 from Google scholar. After duplicate papers were discarded and filters applied a selection of 5,986 studies remain. 5,714 studies were screened out by reading the title paper, abstract and full paper for the following reasons; 5,535 did not

meet the inclusion criteria, 123 studies had a Jadad Scale less than three (3), 35 studies were *in vitro* studies and 33 studies were on diabetic complications. A total of 44 articles were included in this study.

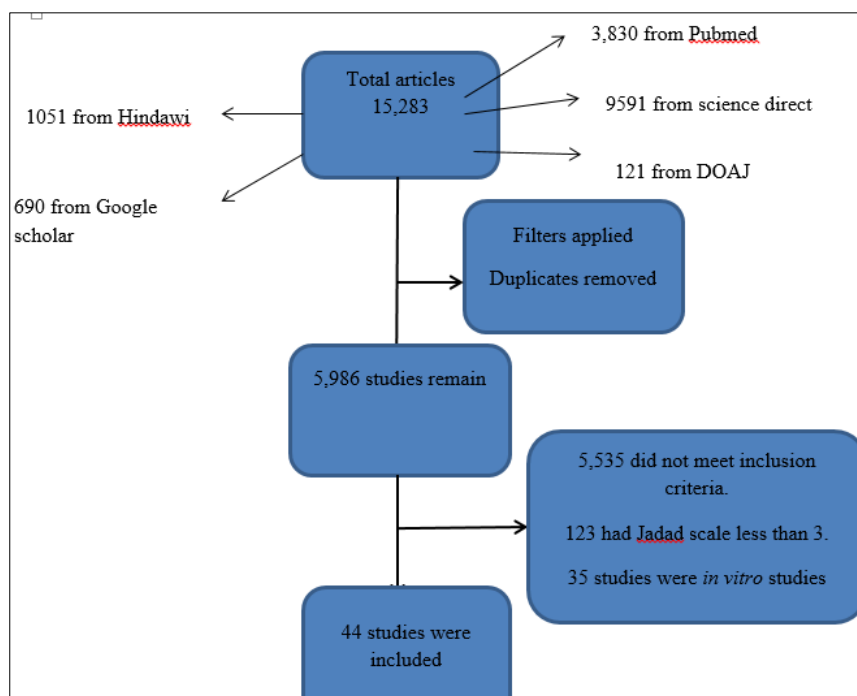


Fig 1: Flowchart of the selection of the articles

Plant parts used as material for the extraction and biological activity should be based on the ethnopharmacological use of the plants. This will direct researchers interested in isolation and characterization of active phytochemicals as well as researchers interested in clinical studies^[57]. In the present study, 35.56% used leaves, 17.78% used stem bark, 11.11% seeds, 4.44% rhizomes, 11.11% fruit, 13.33% roots and 6.67% used mixed plant parts. Among the plants, 60% were trees, 31.11% shrubs and 8.89% herbs.

Chemical methods for the induction of diabetes provide a simple and cost effective model of diabetes in animals^[58]. In the articles reviewed, 55.56% used alloxan to induce diabetes, 42.22% used Streptozocin while 2.22% used combination of Streptozocin and fructose. 88.89% of studies chose the oral route to administer plant extracts, while 11.11% chose the intraperitoneal route.

Conclusion

The systematic review and meta-analysis on Nigerian medicinal plants with antidiabetic activity will provide researchers with valuable information on the most common plants species and methods used.

References

1. World Health Organization. World malaria report 2015. World Health Organization, 2016.
2. Dagogo-Jack, Sam, ed. Diabetes mellitus in developing countries and underserved communities. Springer International Publishing, 2017.

3. Leila M, Rabeta MS, Vikneswaran M, Mohd ZA. Hypoglycemic and anti-hyperglycemic study of *Ocimum tenuiflorum* L. leaves extract in normal and streptozotocin induced diabetic rats. Asian Pac J Trop Biomed. 2016; 6:1029-1036.
4. Chakrabarti R, Rajagopalan R. Diabetes and insulin resistance associated disorders: disease and the therapy. Current science, 2002, 1533-1538.
5. Nwanna EE, Ibukun EO, Oboh G. Inhibitory effects of methanol extracts of two eggplant species from South Western Nigeria on starch hydrolyzing enzymes linked to type 2 diabetes. Afr J Pharm Pharmacol. 2013; 7:1575-84.
6. Ekor M. The growing use of herbal medicines: issues relating to adverse reactions and challenges in monitoring safety. Front Pharmacol. 2014; 4:177.
7. Ezuruike UF, Prieto JM. The use of plants in the traditional management of diabetes in Nigeria: pharmacological and toxicological considerations. J. Ethnopharmacol. 2014; 155(2):857-924.
8. Lee AL, Chen BC, Mou CH, *et al.* Association of traditional Chinese medicine therapy and the risk of vascular complications in patients with type II diabetes mellitus: a nationwide, retrospective, Taiwanese-registry, cohort study. Medicine. 2016; 95(3):e2536.
9. Piero MN, Njagi JM, Kibiti CM, Ngeranwa JJN, Njagi ANM, Njue WM, *et al.* Herbal management of diabetes mellitus: a rapidly expanding research avenue Int J Curr Pharm Res. 2012; 4(2):1-4.

10. Tabatabaei-malazy O, Ramezani A, Atlasi R, Larijani B, Abdollahi M. Scientometric study of academic publications on antioxidative herbal medicines in type 2 diabetes mellitus. *J. Diabetes Metab. Disord.* 2016; 15:48.
11. Moher D, Liberati A, Tetzlaff J, Altman DG, Grp P. Preferred Reporting Items for Systematic Reviews and Meta-Analyses: The PRISMA Statement (Reprinted from *Annals of Internal Medicine*). *Phys. Ther.* 2009; 89:873–880.
12. Jadad, Alejandro R, Henry J. McQuay. "Meta-analyses to evaluate analgesic interventions: a systematic qualitative review of their methodology." *Journal of clinical epidemiology.* 1996; 49.2:235-243.
13. Yerima M, Samaila A. Hypoglycemic and Antioxidant Activity of the Residual Aqueous Extract of *Tamarindus Indica*, *Int. J. Pharm. Research.* 2018; 8(8):69-75.
14. Ochuko L Erukainurea, Olakunle Sannia B, Omamuyovwi M Ijomonec, Collins U Ibejid, Chika I Chukwumaa, Md Shahidul Islam E. The antidiabetic properties of the hot water extract of kola nut (*Cola nitida* (Vent.) Schott & Endl.) in type 2 diabetic rats. *J. of Ethnopharm.* 2019, 242.
15. Innocent Anioke, Chukwugozie Okwuosa, Ikenna Uchendu, Olive Chijioko, Ogechukwu Dozie-Nwakile, Ifeoma Ikegwonu, *et al.* Investigation into Hypoglycemic, Antihyperlipidemic, and Renoprotective Potentials of *Dennettia tripetala* (Pepper Fruit) Seed in a Rat Model of Diabetes; *BioMed Research Int.* 2017, 11. <https://doi.org/10.1155/2017/6923629>
16. Ononamadu CJ, Alhassan AJ, Imam AA, *et al.* In vitro and in vivo evaluation of the antidiabetic and antioxidant activity of methanol extract/ solvent fractions of *ocimum canum* leaves. *Caspian J Intern Med.* 2019; 10(2):162-175.
17. Maxwell Ikechukwu Ezeja, Aruh Ottah Anaga, Isaac U Asuzu. Antidiabetic, antilipidemic, and antioxidant activities of *Gouania longipetala* methanol leaf extract in alloxan-induced diabetic rats, *Pharma. Bio.* 2015; 53(4):605-614. DOI: 10.3109/13880209.2014.935864
18. Folorunsho A Ajayi, Olubukola S Olorunnisola, Adewale Adetutu, Folashade G Olorunfemi, Abiodun O Owoade, Peter Adegbola, *et al.* Anti-hyperglycaemic and Mode of Action of *Thaumatococcus danielli* (BENN.) BENTH Ethanol Leave Extract in Streptozotocin-induced Diabetic Rats; *Asian J. of Res. Med. Pharm. Sci.* 2019; 6(2):1-10.
19. Mhya DH Anigo KM, Umar IA, Alegbejo JO. Evaluation of Hypoglycemic Potential of Extracts of *Balanites Aegyptiaca* Parts; *International Journal of Innov. Res. Adv. Stud.* 2016, 3(9).
20. Ibitoye OB, Uwazie JN, Ajiboye TO. Bioactivity-guided isolation of kaempferol as the antidiabetic principle from *Cucumis sativus* L. fruits. *J Food Biochem.* 2017. <https://doi.org/10.1111/jfbc.12479>.
21. Adeneye AA, Olagunju JA, Olatunji BH, Balogun AF, Akinyele BS, Ayodele MO. Modulatory effect of *Morinda lucida* aqueous stem bark extract on blood glucose and lipid profile in alloxan-induced diabetic rats. *Afr. J. of Biomed. Res.* 2017; 20(1):75-84.
22. Stanley IR Okoduwa, Isamila A Umar, Dorcas B James, Hajiya M Inuwa. Anti-Diabetic Potential of *Ocimum gratissimum* Leaf Fractions in Fortified Diet-Fed Streptozotocin Treated Rat Model of Type-2 Diabetes; *Medicines.* 2017; 4:73.
23. Osasenaga Macdonald Ighodaro, Oluseyi Adeboye Akinloye. Anti-diabetic potential of *Sapium ellipticum* (Hochst) Pax leaf extract in Streptozotocin (STZ)-induced diabetic Wistar rats *BMC Complementary and Alternative Medicine.* 2017; 17:525. DOI 10.1186/s12906-017-2013
24. Ibrahim Fatima Alaba, Lamidi Ajao Usman, Jubril Olayinka Akolade, Oluwafemi Ayodeji Idowu, Azeemat Titilola Abdulazeez, Aliyu Olalekan Amuzat. Antidiabetic Potentials of *Citrus aurantifolia* Leaf Essential Oil. *Thieme,* 2018.
25. Nwodo Okwesili FC, Nweje-Anyalowu Paul C, Joshua Parker E, Uroko Robert. Phytochemical, Antihyperglycaemic and Lipid Profile Effects of Methanol Extract Fraction of *Ricinus communis* Seeds in Alloxan Induced Diabetic Male Wistar Albino. *Asian J. Biochem.* 2016; 11(1):24-33.
26. Joseph L Akpan, Basil C Ezeokpo, Kenneth I Nwadike, Daniel OJ Aja. Antidiabetic and antihyperlipidemic potential of ethanol extract of *Saacia lehmbachii* stem bark in alloxan-induced diabetic rats. *Basic Clin Physiol Pharmacol,* 2018.
27. Jonathan Emeka Emordi, Esther Oluwatoyin Agbaje, Ibrahim Adekunle Oreagba, Osede Ignis Iribhogbe. Antidiabetic Effects of the Ethanolic Root Extract of *Uvaria chamae* P. Beauv (Annonaceae) in Alloxan-Induced Diabetic Rats: A Potential Alternative Treatment for Diabetes Mellitus, *Adv. Pharm. Sci.,* 2018, 13.
28. Sulyman AO, Akolade JO, Sabiu S, Ibrahim RB, Aladodo RA, Ahmad JB, *et al.* Antidiabetic efficacies of methanolic and ethyl acetate extracts of *Aristolochia ringens* (Vahl) roots: in vivo comparative studies. *Com. Clin. Path,* 2019, 1-8.
29. Aina VO, Ibrahim MB, Abdulsalami MS, Adejo GO, Solomon Musa. Anti-Diabetic Activities of the Leaf and Bark Extracts of *Jatropha Curcas* on Alloxan-Induced Diabetic Albino Rats. *J. Nat. Sci. Res.* 2016; 6(4):2016.
30. Emmanuel E Essien, Paul S Thomas. Toxicity, hypoglycemic and antioxidant potentials of *Massularia acuminata* Stem. *J. Pharm. Phytochemistry.* 2018; 7(5):1222-1226.
31. Fageyinbo MS, Akindele AJ, Adenekan SO, Agbaje EO. Evaluation of in-vitro and in-vivo antidiabetic, antilipidemic and antioxidant potentials of aqueous root extract of *Strophanthus hispidus* DC (Apocynaceae). *J. Com. Int. Med.* 2019; 18:16 (3).
32. Idakwoji Precious Adejoh, Okafor Stephen Chiadikaobi, Akuba Ojochegbe Barnabas, Ajayi Oluwakemi Ifeoluwa, Hassan Shehu Muhammed. *In vivo* and *in vitro* comparative evaluation of the anti-diabetic potentials of the parts of *Moringa oleifera* tree *Eur. J. Biotech. Bio.* 2016; 1:14-22.
33. Stanley Irobekhan Reuben Okoduwa1, Ismaila Alhaji Umar, Dorcas Bolanle James, Hajiya Mairo Inuwa. Validation of the Antidiabetic Effects of *Vernonia amygdalina* Delile Leaf Fractions in Fortified Diet-Fed Streptozotocin- Treated Rat Model of Type-2 Diabetes. *J. diabet,* 2019.
34. Omeje EO, Osadebe PO, Obonga WO, Ugwuoke CE. Phytochemical and Pharmacological Basis for the ethnomedicinal use of root extracts from *Anogeissus leiocarpus* as an antidiabetic in Eastern Nigeria. *B. J. Pharm. Res.* 2016; 9(4):1-8.

35. Abubakar MK, Wasagu RS, Usman JN, Galadima LG. Effect of Methanol and Aqueous Leaf Extract of *Mitracarpus Scabrum* in Alloxan Induced Diabetic Rats. *J. Pharmacog. Phytochem.* 2016; 1, 5(1):4.
36. Jubril Olayinka Akolade, Lamidi Ajao Usman, Omoaruemike Ebele Okereke, Nasir Olarewaju Muhammad. Antidiabetic Potentials of Essential Oil Extracted. *J Med Food.* 2017; 17(10):1122-1128.
37. Mohammed Auwal brahim, M Shahidul Islam. Anti-diabetic effects of the acetone fraction of *Senna singueana* stem bark in a type 2 diabetes rat model. *J. Ethnopharmacol.* 2014; 153:392–399.
38. Mohammed Auwal Ibrahim, James Dama Habilac, Neil Anthony Koorbanally, Md Shahidul Islam. Butanol fraction of *Parkia biglobosa* (Jacq.) G. Don leaves enhance β -cell functions, stimulates insulin secretion and ameliorates other type 2 diabetes-associated complications in rats. *J. Ethnopharm.*
39. Oloyede OB, Ajiboye TO, Abdussalam AF, Adeleye AO. *Blighia sapida* leaves halt elevated blood glucose, dyslipidemia and oxidative stress in alloxan-induced diabetic rats. *J. Ethnopharmacol.* 2014. <http://dx.doi.org/10.1016/j.jep.2014.08.022i>
40. Engwa Azeh Godwill, Marcellus Unaegbu, Aniakor Uchenna Esther, Osuji Amarachukwu Gloria, Agbafor Nwonu Kingsley, Olayinka Ayobami Aiyegoro, et al. Antioxidant and antidiabetic activities of the seed and leaf extracts of *Chrysophyllum albidum*, 2016. doi: 10.1016/S2222-1808(16)61102-7.
41. Anyanwu GO, Iqbal J, Khan SU, Zaib S, Rauf K, Onyeneke CE, et al. Antidiabetic activities of chloroform fraction of *Anthocleista vogelii* Planch root bark in rats with diet-and alloxan-induced obesity-diabetes. *J. ethnopharm.* 2019; 229:293-302.
42. Obafemi T Olabisi, Olaleye Tolulope M, Afolabi C Akinmoladun. Antidiabetic property of miracle fruit plant (*Synsepalum dulcificum* Shumach. & Thonn. Daniell) leaf extracts in fructose-fed streptozotocin-injected rats via anti-inflammatory activity and inhibition of carbohydrate metabolizing enzymes, 2019.
43. Ochuko L Erukainurea, Olakunle Sannia B, Omamuyovwi M Ijomonec, Collins U Ibejid, Chika I Chukwumaa, Md Shahidul Islam E. The antidiabetic properties of the hot water extract of kola nut *Cola nitida* (Vent.) Schott & Endl.) in type 2 diabetic rats. *J. Ethnopharm.* 2019, 242.
44. Kayode Olayele Karigidi, Charles Ojo Olaiya. Antidiabetic activity of corn steep liquor extract of *Curculigo pilosa* and its solvent fractions in streptozotocin-induced diabetic rats. (Apocynaceae). *J. Com. Int. Med.* 2019, 20180055.
45. Nadro MS, Elkanah G. Hypoglycaemic effect of fractions and crude methanolic leaf extract of *Phyllanthus fraternus* in streptozotocin -induced diabetic and normal rats. *J. Med. Plants Res.* 2017, 60.
46. Abiola T, Dibie DC, Akinwale OJ, Shomuyiwa OA. Assessment of the antidiabetic potential of the ethanolic extract of date palm (*Phoenix Dactylifera*) seed in alloxan-induced diabetic rats. *J. Diabet and Met.* 2018; 9(784):1-9.
47. Arise O Rotimi, Tosan Akapa, Moshood A Adigun, Abeebe A Yekeen, Oluwafemi O Oguntibeju. Normoglycaemic, Normolipidaemic and Antioxidant Effects of Ethanolic Extract of *Acacia ataxacantha* Root in Streptozotocin -induced Diabetic Rats. *Not Sci Biol.* 2016; 8(2):144-150. DOI: 10.15835/nsb.8.2.9770.
48. Issaka J, Larbie C, Tandoh MA. Anti-diabetic effect of aqueous fruit extract of *Borassus aethiopum* (Mart.) in alloxan-induced diabetic rats. *Int. J. Phytomed.* 2016; 8(3):384-397.
49. Achi NK, Ohaeri OC, Ijeh II, Eleazu C. Modulation of the lipid profile and insulin levels of streptozotocin induced diabetic rats by ethanol extract of *Cnidioscolus aconitifolius* leaves and some fractions: Effect on the oral glucose tolerance of normoglycemic rats. *Biomed. Pharmacotherapy.* 2017; 86:562–569.
50. Muhammad U, Jarumi IK, Alhassan AJ, Wudil AM, Dangambo MA. Acute Toxicity and Hypoglycemic Activity of Aqueous Fruit Pulp Extract of *Adansonia digitata* L. (Afpead) on Alloxan Induced Diabetic Rats. *J. Adv. Med. Pharm. Sci.* 2016; 6(3):1-6.
51. Osigwe CC, Akah PA, Nworu CS, Okoye TC, Tchimine MK. Antihyperglycemic studies on the leaf extract and active fractions of *Newbouldia laevis* (Bignoniaceae). *Pharmacology & Pharmacy.* 2015; 6(11):518.
52. Odoh UE, Onugha VO, Chukwube VO. Evaluation of antidiabetic effect and hematological profile of methanol extract of *Ceiba pentandra* G (Malvaceae) stem bark on alloxan-induced diabetic rats. *Afr. J. Pharm. Pharmacol.* 2016; 10(28):584-90.
53. Osamuyi H Uwumarongie, Nkem D Onwukaeme, Ighodaro Igbe. Phytochemical Analysis and Hypoglycaemic Activity of the Methanol Stem Bark Extract of *Tabernaemontana pachysiphon* Stapf (Apocynaceae). *Trop J Nat Prod Res.* 2017; 1(5):213-216.
54. Agbafor KN, Onuohah SC, Ominyi MC, Orinya OF, Ezeani N, Alum E. Antidiabetic, Hypolipidemic and Antiathrogenic Properties of Leaf Extracts of *Ageratum conyzoides* in Streptozotocin-Induced diabetic rats. *Int. J. Curr. Microbiol. App. Sci.* 2015; 4(11):816-824.
55. Dickson RA, Harley BK, Berkoh D, Ngala RA, Titiloye NA, Fleischer TC. Antidiabetic and Haematological Effect of *Myrianthus arboreus* P. Beauv. Stem Bark Extract in Streptozotocin - Induced Diabetic rats. *IJPSR.* 2016; 7:12.
56. Oloyede HOB, Bello TO, Ajiboye TO, Salawu MO. Antidiabetic and antidyslipidemic activities of aqueous leaf extract of *Dioscoreophyllum cumminsii* (Stapf) Diels in alloxan-induced diabetic rats. *J. Ethnopharm.* 2015; 166:313–322.
57. Munhoz CM, Antonio and Frode S Tania. Isolated Compounds from Natural Products with Potential Antidiabetic Activity - A Systematic Review. *Curr. Diabet. Rev.* 2018; 14:36-106.
58. King AJ. The use of animal models in diabetes research: Animal models of diabetes. *Br J Pharmacol.* 2012; 166(3):877-94.